



Stereoselective synthesis of (2*R*,3*R*,4*R*)-3-hydroxy-2,4,6-trimethylheptanoic acid and determination of the absolute stereochemistry of the natural product from callipeltin A

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Received 3 June 2002; accepted 20 June 2002

Abstract—A revised stereostructure of 3-hydroxy-2,4,6-trimethylheptanoic acid, the β -hydroxy acid that acylates the N-terminus of callipeltin A, is proposed on the basis of analysis of *J*-coupling in the ^1H NMR spectrum of the acetonide derivative obtained from the acid hydrolysate of callipeltin A. The proposed structure was definitively confirmed by enantioselective synthesis. © 2002 Elsevier Science Ltd. All rights reserved.

Recently, we reported the synthesis of (2*R*,3*R*,4*S*)-3-hydroxy-2,4,6-trimethylheptanoic acid, the proposed β -hydroxy acid unit that acylates the N-terminus of callipeltin A **1**.¹ However, we found that its ^1H NMR spectral data were not identical with those of the corresponding unit obtained from the acid hydrolysate of callipeltin A **1**, thus indicating that the absolute configuration of the aforementioned natural unit should be revised. Because the (3*R*)-stereochemistry at the carbinol centre was unambiguously established by a modified Mosher's method,² the stereochemical revision should concern the C(2) stereogenic centre, the C(4) stereogenic centre, or both.

In order to gain more insights into the stereochemistry of the natural β -hydroxy acid unit, we subjected a 40 mg sample of callipeltin A **1** to acid hydrolysis followed by dichloromethane extraction, to obtain 4.0 mg of the natural β -hydroxy acid **2**. This latter was readily transformed in the 1,3-*O*-isopropylidene derivative **4** by the three steps sequence depicted in the Scheme 1. The vicinal coupling constant for H2/H3 (8.2 Hz) was indicative of an *anti*-relationship between these two

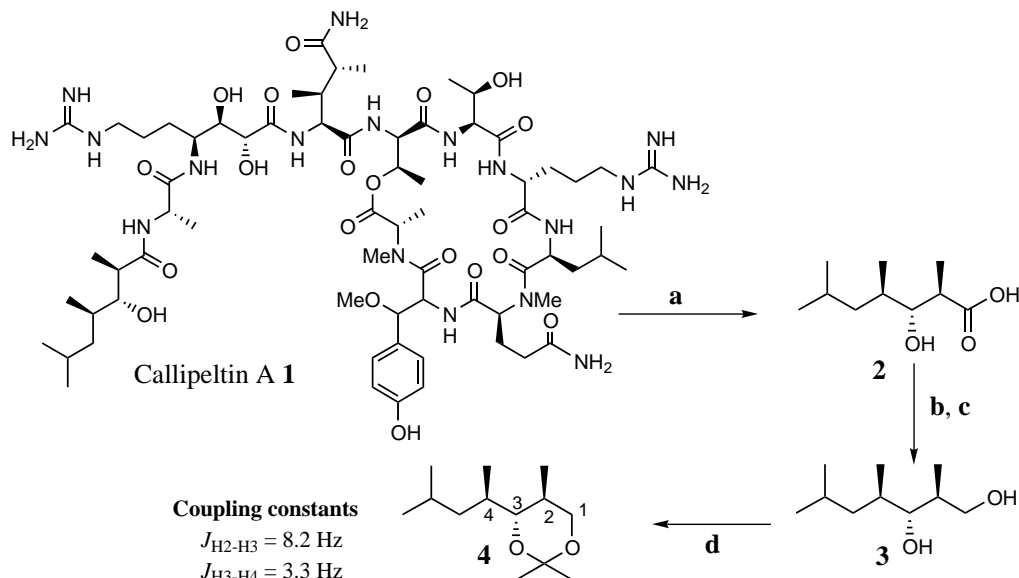
protons.³ Thus, the (2*R*,3*R*) configuration was firmly established.

From these results we reached the conclusion that the correct stereochemistry of the natural β -hydroxy acid **2** must be (2*R*,3*R*,4*R*). This revised stereochemistry has been confirmed by enantioselective synthesis as described herein.

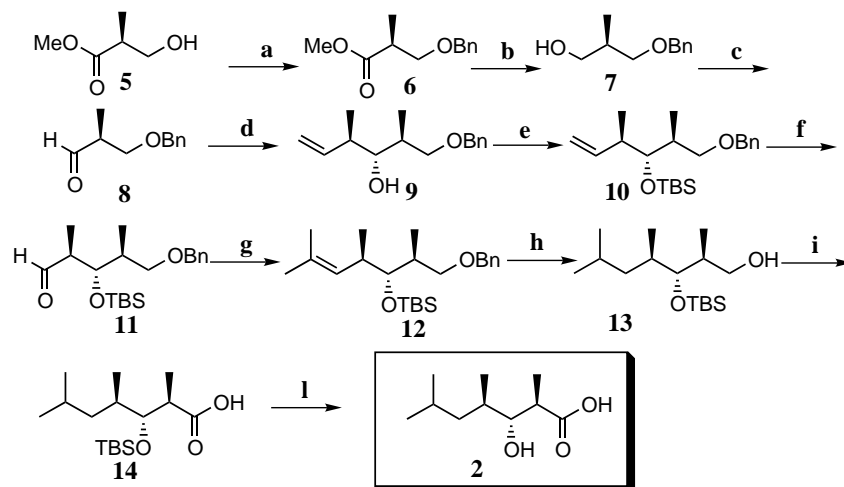
Our planned synthetic route to the above fragment **2**, outlined in Scheme 2, is similar to that used for the synthesis of (2*R*,3*R*,4*S*)-3-hydroxy-2,4,6-trimethylheptanoic acid.¹ Indeed we elaborated a flexible synthetic strategy which was amenable to the construction of the different stereoisomers of the β -hydroxy acid unit in **1**.

In this case Brown's crotylboration⁴ was selected as the key step for the assembly of the C3–C4 *anti*-propionate unit in β -hydroxy acid **2**. Thus, aldehyde **8**, obtained in high yield (three steps, 87%) from commercially available methyl (2*S*)-2-methyl-3-hydroxy propionate **5**, was reacted with the allyl borane derived from (+)-*B*-methoxydiisopinocampheyl borane and (*E*)-butene to give the *anti*-homoallylic alcohol **9**. The diastereomeric purity of **9** was >98% as judged by the NMR spectra of the crude reaction mixture. We encountered difficulties in separating **9** in acceptable yield from the isopinocampheol arising from the reaction. Thus, crude **9** was converted into the TBS-protected derivative **10** which was easily separated from the protected isopinocampheol.

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Scheme 1. Reagents and conditions: (a) HCl 6N, 130°C, 6 h, then CH₂Cl₂ extraction; (b) CH₂N₂, rt, quantitative; (c) LiBH₄, dry MeOH, 2 h, 96%; (d) dimethoxypropane dry, *p*-TsOH (cat.), quantitative.



Scheme 2. Reagents and conditions: (a) BTCA, TfOH, 0°C to rt, 3 h, 94%; (b) LiBH₄, dry MeOH, 2 h, 96%; (c) DMSO, (COCl₂), -78°C, 30 min, then 7, Et₃N, -78°C to rt, 1 h, 97%; (d) *t*-BuOK, (*E*)-but-2-ene, *n*-BuLi, -78 to -45°C, (+)-*B*-methoxydiisopinocampheylborane, BF₃·OEt₂, aldehyde 8, -78°C, 4 h; (e) 2,6-lutidine, TBSOTf, 0°C to rt, 2 h, 76%, two steps; (f) OsO₄/*N*-methylmorpholine-*N*-oxide, rt, 12 h, then H₅IO₆ 15 min, 97%; (g) isopropyltriphenylphosphonium iodide, *n*-BuLi, rt 3 h, 84%; (h) H₂/Pd(OH)₂, 3 atm, 2 days, 90%; (i) NaIO₄/RuCl₃·H₂O (cat.), 97%; (l) MeOH/HCl, 25°C, 3 h, quantitative.

pheol by silica gel chromatography (76% yield of 10 from 8).⁵ Oxidative cleavage of the terminal double bond (OsO₄, NMO, acetone–water, H₅IO₆) led to the aldehyde 11. This latter was subjected to the same reaction sequence previously developed by us for the synthesis of (2*R*,3*R*,4*S*)-3-hydroxy-2,4,6-trimethylheptanoic acid.¹ Thus, Wittig olefination followed by one-pot reduction of the alkene bond and removal of the benzyl protecting group, afforded alcohol 13 which was smoothly oxidised to the protected β-hydroxy acid 14.

The complete overlap of the NMR data of the synthetic derivative 2,⁶ obtained by acid hydrolysis of 14 (MeOH/HCl), with that of the corresponding natural

unit 2, unambiguously defined the (2*R*,3*R*,4*R*)-configuration. Further confirmation of the absolute stereochemistry of natural 2 was obtained from comparison of specific rotation data (synthetic 2: [α]_D = +14.9, (*c* 0.4), CHCl₃; natural 2: [α]_D = +14.4, (*c* 0.4, CHCl₃).

In conclusion, the stereochemistry of the β-hydroxy acid unit that acylates the N-terminus of the cyclodeptide callipeltin A 1 was definitively established by asymmetric synthesis. Furthermore, our synthesis of the protected β-hydroxy acid 14 is short (nine steps from the commercially available L-malate dimethyl ester 5), efficient (47% overall yield) and amenable to scale-up.

Acknowledgements

This work was supported by grants from MURST (PRIN 2001) 'Sostanze naturali ed analoghi sintetici ad attività antitumorale' Rome, Italy. Mass spectra were provided by the CRIAS Centro Interdipartimentale di Analisi Strumentale, Faculty of Pharmacy, University of Naples. We acknowledge the ungraduated student Luigi Fontana for his kind assistance and the staff are also acknowledged. The NMR spectra were recorded at CRIAS Centro Interdipartimentale di Analisi Strumentale, Faculty of Pharmacy, University of Naples.

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3. ^1H NMR (CDCl_3 , 500 MHz) for compound **4**, δ : 4.22 (1H, dd, $J=10.3$, 5.1 Hz, H-1a), 3.75 (1H, dd, $J=10.3$, 2.7 Hz, H-1b), 3.40 (1H, dd, $J=8.2$, 3.3 Hz, H-3), 1.62 (1H, m, H-4), 1.60 (2H, m's, H-2, H-6), 1.30/1.32 (6H, s's, acetone Me), 1.20 (1H, m, H-5a), 1.13 (1H, m, H-5b), 0.94 (3H, d, $J=6.6$ Hz, Me-8), 0.90 (3H, d, $J=6.6$ Hz, Me-9), 0.86 (3H, d, $J=6.6$ Hz, Me-10), 0.84 (3H, d, $J=6.6$ Hz, Me-7).
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5. NMR data (CDCl_3 , 500 MHz) and specific rotation (chloroform) for compound **10**. ^1H NMR δ : 7.34 (5H, m, *benzyl*), 5.95 (1H, m, H-5), 5.03 (1H, d, $J=5.2$, H-6a), 5.0 (1H, s, H-6b), 4.50 (2H, s, benzylic CH_2), 3.60 (2H, m, H-1a, H-3), 3.40 (1H, t, $J=8.8$ Hz, H-1b), 2.41 (1H, m, H-4), 2.01 (1H, m, H-2) 1.08 (3H, d, $J=6.6$ Hz, Me-4), 1.02 (3H, d, $J=6.6$ Hz, Me-2), 0.95 (9H, s, Si- $\text{C}(\text{CH}_3)_3$), 0.1 (3H, s, Si- $\text{C}(\text{CH}_3)_3$), 0.12 (3H, s, Si- $\text{C}(\text{CH}_3)_3$); ^{13}C NMR δ : 141.3, 138.8, 128.3, 127.6, 127.4, 115.1, 79.6, 77.8, 73.0, 72.9, 42.0, 38.2, 26.1, 18.4, 18.2, 14.9, -3.8, -4.0; $[\alpha]_{\text{D}} = -4.3$ (c 2.2, CHCl_3).
6. NMR data (CDCl_3 , 500 MHz) and specific rotation (chloroform) for synthetic **2**. ^1H NMR δ : 3.48 (1H, dd, $J=7.7$, 4.1 Hz, H-3), 2.73 (1H, q, $J=7.7$ Hz, H-2), 1.70 (1H, m, H-4), 1.64 (1H, m, H-6), 1.26 (3H, d, $J=7.7$ Hz, Me-2), 1.18 (1H, m, H-5a), 1.12 (1H, m, H-5b), 0.96 (3H, d, $J=5.8$ Hz, Me-6), 0.92 (3H, d, $J=6.8$ Hz, Me-6), 0.84 (3H, d, $J=6.8$ Hz, Me-4); ^{13}C NMR δ : 180.7, 78.2, 42.1, 39.4, 33.4, 25.2, 24.2, 21.2, 16.6, 14.6; $[\alpha]_{\text{D}} = +14.9$ (c 0.4, CHCl_3).